**Amendments to the Specification:**

Please replace the paragraph beginning at page 32, line 17, with the following:

--RNA was isolated for both Northern blotting and quantitative real-time PCR (Q-PCR) using a standard guanidium isothiocyanate method. Northern blotting was done as previously described (Lee, H., Dadgostar, H., Cheng, Q., Shu, J., and Cheng, G. (1999), Proc. Natl. Acad. Sci. USA *96*, 9136-9141), and was hybridized using a RANTES cDNA fragment (IMAGE Clone: 832342, Research Genetics). For Q-PCR, RNA was quantitated and 2$\mu$g of RNA was used to make cDNA templates using Superscript II (Invitrogen) according ~~the the manufactuors instruction~~ to the manufacturer's instructions with either oligo-dT or random hexamer as primers. Q-PCR analyses ~~was~~ were done using the iCycler thermocycler (Bio-Rad). Q-PCR was conducted in a final volume of 25$\mu$L containing: Taq polymerase, lx Taq buffer (Stratagene), 125 $\mu$M dNTP, SYBR™ Green I (Molecular Probes), and ~~Fluoroscein~~ Fluorescein (Bio-Rad), using oligo-dT cDNA or random hexamer cDNA as the PCR template. Amplification conditions were: 95°C (3 min), 40 cycles of 95°C (20 sec), 55°C (30 sec), 72°C (20 sec). The following primers were used to amplify a specific 100-120 bp fragment of the following genes:

RANTES 5': GCCCACGTCAAGGAGTATTTCTA (SEQ ID NO:1),

RANTES 3': ACACACTTGGCGGTTCCTTC (SEQ ID NO:2),

Mxl 5': AAACCTGATCCGACTTCACTTCC (SEQ ID NO:3),

Mxl 3': TGATCGTCTTCAAGGTTTCCTTGT (SEQ ID NO:4),

IFI1 5': CCAGAGCATGGGAAAGAGGTT (SEQ ID NO:5),

IFI1 3': CCGGACCTCTGATAGGACACTG (SEQ ID NO:6),

IFI-204 5': TTGGCTGCAATGGGTTCAT (SEQ ID NO:7),

IFI-204 3': AGT GGGATATTCATTGGTTCGC (SEQ ID NO:8),

IRF7 5': ACAGGGCGTTTTATCTTGCG (SEQ ID NO:9),

IRF7 3': TCCAAGCTCCCGGCT AAGT (SEQ ID NO:10),

IP-10 5': CCTGCCCACGTGTTGAGAT (SEQ ID NO:11),

IP-10 3': TGATGGTCTTAGATTCCGGATTC (SEQ ID NO:12),

ISG-15 5': CAGGACGGTCTTACCCTTT CC (SEQ ID NO:13),

ISG-15 3': AGGCTCGCTGCAGTTCTGTAC (SEQ ID NO:14),

IFIT1 5': GGCAGGAACAATGTGCAAGAA (SEQ ID NO:15),

IFIT1 3': CTCAAATGTGGGCCTCAGTT (SEQ ID NO:16),

18S 5': CCGCGGTTCTATTTTGTTGGT (SEQ ID NO:17),

18S 3': CTCTAGCGGCGCAATACGA (SEQ ID NO:18),

IFN-$\beta$ 5': AGCTCCAAGAAAGGACGAACAT (SEQ ID NO:19),

IFN-$\beta$ 3': GCCCTGTAGGTGAGGTTGATCT (SEQ ID NO:20),

I$\kappa$B$\alpha$ 5': CTGCAGGCCACCAACTACAA (SEQ ID NO:21),

I$\kappa$B$\alpha$ 3': CAGCACCCAAAGTCACCAAGT (SEQ ID NO:22),

Beta Actin 5': AGGTGTGCACCTTTTATTGGTCTCAA (SEQ ID NO:23),

Beta Actin 3': TGTATGAAGGTTTGGTCTCCCT (SEQ ID NO:24).--

Please replace the paragraph beginning at page 33, line 28, with the following:

--The full-length and dominant-negative IRF3-expression plasmids were created by PCR ~~amplifiction~~ amplification of IRF3 cDNA (IMAGE clone: 3666172) using either IRF3(1-420) 5'-CAGGACTGATCAACCATGGAAACCCCGAAACCGCGGATT-3' (SEQ ID NO:25) or IRF3DBD(133-420) 5'-CAGGACATCCATGCACTCCCAGGAAAACCTACCGAAG-3' (SEQ ID NO:26) in conjunction with the 3' primer 5'-CAGGACGCGGCCGCGATATTCCAGT GGCCTGGAAGTC-3' (SEQ ID NO:27). Fragments were cloned into the ~~BglI/NotI~~ BglI/NotI

orBamHI/NotI sites of pEBB-puro. pCDNA3-IkBm-ER was constructed as described (Lee, H., Dadgostar, H., Cheng, Q., Shu, J., and Cheng, G. (1999), Proc. Natl. Acad. Sci. USA *96*, 9136-9141). The –243 IP10 pCAT plasmid was a kind gift of Thomas A. Hamilton.--

Please replace the paragraph beginning at page 47, line 7, with the following:

--RNA was isolated by standard guanidium isothiocyanate methods. cDNA template for quantitative realtime PCR analysis was then synthesized and PCR was performed using the iCycler thermocycler (Bio-Rad) as previously described (Doyle, S. E., S. A. Vaidya, R. O'Connell, H. Dadgostar, P. W. Dempsey, T.-T. Wu, G. Rao, R. Sun, M. E. Haberland, R. L. Modlin, and G. Cheng. 2002. IRF3 mediates a TLR3/TLR4-specific antiviral gene program. *Immunity 17:251*). IFN-βIP10, IκBα and IFI-204 primers were the same as those previously described (Doyle, S. E., S. A. Vaidya, R. O'Connell, H. Dadgostar, P. W. Dempsey, T.-T. Wu, G. Rao, R. Sun, M. E. Haberland, R. L. Modlin, and G. Cheng. 2002. ~~IRF3 mediates a TLR3/TLR4-specific antiviral gene program. *Immunity 17:251*).~~ For other genes, the following primers were used:

IL-6 Forward: CACAGAGGATACCACTCCCAACA (SEQ ID NO:28) and

Reverse: TCCACGATTTCCCAGAGAACA (SEQ ID NO:29);

TLR3 Forward: TCTGGAAACGCGCAAACC (SEQ ID NO:30) and

Reverse: GCCGTTGGACTCTAAATTCAAGAT (SEQ ID NO:31);

TLR4 Forward: AGAAATTCCTGCAGTGGGTCA (SEQ ID NO:32) and

Reverse: TCTCTACAGGTGTTGCACATGTCA (SEQ ID NO:33);

TIRAP Forward: CAGGCAGGCTCTGTTGAAGAA (SEQ ID NO:34) and

Reverse: TGTGTGGCTGTCTGTGAACCA (SEQ ID NO:35);

MyD88 Forward: CATGGTGGTGGTTGTTTCTGAC (SEQ ID NO:36) and

Reverse: TGGAGACAGGCTGAGTGCAA (SEQ ID NO:37); and

ICAM1 Forward: TGTCAGCCACTGCCTTGGTA (SEQ ID NO:38) and

Reverse: CAGGATCTGGTCCGCTAGCT[[.]] SEQ ID NO:39);

L32 Forward: AAGCGAAACTGGCGGAAAC (SEQ ID NO:40) and

Reverse: TAACCGATGTTGGGCATCAG (SEQ ID NO:41).--

Please insert the accompanying paper copy of the Sequence Listing, page numbers 1 to 10, at the end of the application.